

Ref #	Hits	Search Query	DBs	Default Operator	Plurals	Time Stamp
L1	111020	(synthetic or variant or modif\$ or alter\$) near5 (gene\$1 or sequence\$1 or nucleic acid\$1)	US-PGPUB; USPAT	ADJ	OFF	2004/08/25 16:19
L3	4327	codon near3 (choice\$1 or preference\$1 or select\$)	US-PGPUB; USPAT	ADJ	OFF	2004/08/25 16:21
L4	4126	1 and 3	US-PGPUB; USPAT	ADJ	OFF	2004/08/25 16:22
L5	2180	1 same 3	US-PGPUB; USPAT	ADJ	OFF	2004/08/25 16:27
L6	49028	(transcription factor\$1 or splice or promoter\$1 or polyadenylat\$) near5 (site\$1 or sequence\$1)	US-PGPUB; USPAT	ADJ	OFF	2004/08/25 16:26
L7	14346	1 same 6	US-PGPUB; USPAT	ADJ	OFF	2004/08/25 16:27
L8	1251	5 and 7	US-PGPUB; USPAT	ADJ	OFF	2004/08/25 16:27
L9	134	1 same 3 same 6	US-PGPUB; USPAT	ADJ	OFF	2004/08/25 16:28

* * * * * * * * * * * * * * * STN Columbus * * * * * * * * * * * * * * *

FILE 'HOME' ENTERED AT 10:06:28 ON 25 AUG 2004

=> fil.bec
COST IN U.S. DOLLARS
FULL ESTIMATED COST

SINCE FILE ENTRY TOTAL
SESSION 0.21 0.21

FILES 'MEDLINE, SCISEARCH, LIFESCI, BIOTECHDS, BIOSIS, EMBASE, HCAPLUS, NTIS,
ESBIOBASE, BIOTECHNO, WPIDS' ENTERED AT 10:06:48 ON 25 AUG 2004
ALL COPYRIGHTS AND RESTRICTIONS APPLY. SEE HELP USAGETERMS FOR DETAILS.

11 FILES IN THE FILE LIST

=> s (synthetic or variant or modif? or alter?) (5a) (gene/q or nucleic acid#)
FILE 'MEDLINE'

121539 SYNTHETIC
58643 VARIANT
359543 MODIF?
631953 ALTER?
170110 NUCLEIC
1478431 ACID#
169729 NUCLEIC ACID#
(NUCLEIC(W)ACID#)
L1 40974 (SYNTHETIC OR VARIANT OR MODIF? OR ALTER?) (5A) (GENE/Q OR NUCLEIC
ACID#)

FILE 'SCISEARCH'

150092 SYNTHETIC
62205 VARIANT
472532 MODIF?
621439 ALTER?
31869 NUCLEIC
1174612 ACID#
31406 NUCLEIC ACID#
(NUCLEIC(W)ACID#)
L2 40575 (SYNTHETIC OR VARIANT OR MODIF? OR ALTER?) (5A) (GENE/Q OR NUCLEIC
ACID#)

FILE 'LIFESCI'

38384 SYNTHETIC
17702 VARIANT
91970 MODIF?
171688 ALTER?
12540 "NUCLEIC"
311433 ACID#
12381 NUCLEIC ACID#
(NUCLEIC(W)ACID#)
L3 21080 (SYNTHETIC OR VARIANT OR MODIF? OR ALTER?) (5A) (GENE/Q OR NUCLEIC
ACID#)

FILE 'BIOTECHDS'

12453 SYNTHETIC
8338 VARIANT
31626 MODIF?
24266 ALTER?
37631 NUCLEIC
127373 ACID#
37553 NUCLEIC ACID#
(NUCLEIC(W)ACID#)
L4 13943 (SYNTHETIC OR VARIANT OR MODIF? OR ALTER?) (5A) (GENE/Q OR NUCLEIC
ACID#)

FILE 'BIOSIS'
 191612 SYNTHETIC
 59943 VARIANT
 361830 MODIF?
 640412 ALTER?
 49238 NUCLEIC
 1306370 ACID#
 48659 NUCLEIC ACID#
 (NUCLEIC(W)ACID#)
L5 46345 (SYNTHETIC OR VARIANT OR MODIF? OR ALTER?) (5A) (GENE/Q OR NUCLEIC
 ACID#)

FILE 'EMBASE'
 103573 SYNTHETIC
 54345 VARIANT
 322451 MODIF?
 593897 ALTER?
 32902 "NUCLEIC"
 1285302 ACID#
 32621 NUCLEIC ACID#
 ("NUCLEIC"(W)ACID#)
L6 37544 (SYNTHETIC OR VARIANT OR MODIF? OR ALTER?) (5A) (GENE/Q OR NUCLEIC
 ACID#)

FILE 'HCAPLUS'
 542134 SYNTHETIC
 54542 VARIANT
 875783 MODIF?
 800345 ALTER?
 159274 NUCLEIC
 4330304 ACID#
 158327 NUCLEIC ACID#
 (NUCLEIC(W)ACID#)
L7 66631 (SYNTHETIC OR VARIANT OR MODIF? OR ALTER?) (5A) (GENE/Q OR NUCLEIC
 ACID#)

FILE 'NTIS'
 18688 SYNTHETIC
 2516 VARIANT
 96083 MODIF?
 89855 ALTER?
 1798 NUCLEIC
 54223 ACID#
 1782 NUCLEIC ACID#
 (NUCLEIC(W)ACID#)
L8 931 (SYNTHETIC OR VARIANT OR MODIF? OR ALTER?) (5A) (GENE/Q OR NUCLEIC
 ACID#)

FILE 'ESBIOBASE'
 37324 SYNTHETIC
 22490 VARIANT
 134896 MODIF?
 218291 ALTER?
 23250 NUCLEIC
 340126 ACID#
 23138 NUCLEIC ACID#
 (NUCLEIC(W)ACID#)
L9 22511 (SYNTHETIC OR VARIANT OR MODIF? OR ALTER?) (5A) (GENE/Q OR NUCLEIC
 ACID#)

FILE 'BIOTECHNO'
 41250 SYNTHETIC
 25068 VARIANT
 86734 MODIF?

148127 ALTER?
19939 NUCLEIC
371908 ACID#
19837 NUCLEIC ACID#
(NUCLEIC(W)ACID#)
L10 27490 (SYNTHETIC OR VARIANT OR MODIF? OR ALTER?) (5A) (GENE/Q OR NUCLEIC
ACID#)

FILE 'WPIDS'
209332 SYNTHETIC
17461 VARIANT
255063 MODIF?
415925 ALTER?
49792 NUCLEIC
897346 ACID#
49543 NUCLEIC ACID#
(NUCLEIC(W)ACID#)
L11 16597 (SYNTHETIC OR VARIANT OR MODIF? OR ALTER?) (5A) (GENE/Q OR NUCLEIC
ACID#)

TOTAL FOR ALL FILES
L12 334621 (SYNTHETIC OR VARIANT OR MODIF? OR ALTER?) (5A) (GENE/Q OR NUCLEIC
ACID#)

=> s codon(3a) (choice# or preference# or select?)
FILE 'MEDLINE'
33513 CODON
120064 CHOICE#
43909 PREFERENCE#
629127 SELECT?
L13 429 CODON (3A) (CHOICE# OR PREFERENCE# OR SELECT?)

FILE 'SCISEARCH'
23775 CODON
114362 CHOICE#
58296 PREFERENCE#
798493 SELECT?
L14 421 CODON (3A) (CHOICE# OR PREFERENCE# OR SELECT?)

FILE 'LIFESCI'
13885 CODON
19544 CHOICE#
27603 PREFERENCE#
202991 SELECT?
L15 311 CODON (3A) (CHOICE# OR PREFERENCE# OR SELECT?)

FILE 'BIOTECHDS'
4791 CODON
1425 CHOICE#
802 PREFERENCE#
59459 SELECT?
L16 104 CODON (3A) (CHOICE# OR PREFERENCE# OR SELECT?)

FILE 'BIOSIS'
27810 CODON
74411 CHOICE#
59362 PREFERENCE#
682423 SELECT?
L17 468 CODON (3A) (CHOICE# OR PREFERENCE# OR SELECT?)

FILE 'EMBASE'
26626 CODON
105876 CHOICE#
37145 PREFERENCE#

576300 SELECT?
L18 368 CODON (3A) (CHOICE# OR PREFERENCE# OR SELECT?)

FILE 'HCAPLUS'
 32831 CODON
 79749 CHOICE#
 40260 PREFERENCE#
 1102568 SELECT?
L19 649 CODON (3A) (CHOICE# OR PREFERENCE# OR SELECT?)

FILE 'NTIS'
 90 CODON
 19166 CHOICE#
 4813 PREFERENCE#
 162925 SELECT?
L20 2 CODON (3A) (CHOICE# OR PREFERENCE# OR SELECT?)

FILE 'ESBIOBASE'
 13569 CODON
 30746 CHOICE#
 18380 PREFERENCE#
 245235 SELECT?
L21 236 CODON (3A) (CHOICE# OR PREFERENCE# OR SELECT?)

FILE 'BIOTECHNO'
 21971 CODON
 8409 CHOICE#
 7785 PREFERENCE#
 148138 SELECT?
L22 314 CODON (3A) (CHOICE# OR PREFERENCE# OR SELECT?)

FILE 'WPIDS'
 2513 CODON
 26192 CHOICE#
 6230 PREFERENCE#
 987036 SELECT?
L23 73 CODON (3A) (CHOICE# OR PREFERENCE# OR SELECT?)

TOTAL FOR ALL FILES
L24 3375 CODON (3A) (CHOICE# OR PREFERENCE# OR SELECT?)

=> S 112 and 124

FILE 'MEDLINE'
L25 36 L1 AND L13

FILE 'SCISEARCH'
L26 23 L2 AND L14

FILE 'LIFESCI'
L27 18 L3 AND L15

FILE 'BIOTECHDS'
L28 31 L4 AND L16

FILE 'BIOSIS'
L29 31 L5 AND L17

FILE 'EMBASE'
L30 24 L6 AND L18

FILE 'HCAPLUS'
L31 73 L7 AND L19

FILE 'NTIS'

L32 0 L8 AND L20

FILE 'ESBIOBASE'

L33 15 L9 AND L21

FILE 'BIOTECHNO'

L34 20 L10 AND L22

FILE 'WPIDS'

L35 22 L11 AND L23

TOTAL FOR ALL FILES

L36 293 L12 AND L24

=> s l36 not 2001-2004/py

FILE 'MEDLINE'

1967013 2001-2004/PY

L37 30 L25 NOT 2001-2004/PY

FILE 'SCISEARCH'

3631421 2001-2004/PY

L38 19 L26 NOT 2001-2004/PY

FILE 'LIFESCI'

359020 2001-2004/PY

L39 16 L27 NOT 2001-2004/PY

FILE 'BIOTECHDS'

76520 2001-2004/PY

L40 14 L28 NOT 2001-2004/PY

FILE 'BIOSIS'

1871918 2001-2004/PY

L41 28 L29 NOT 2001-2004/PY

FILE 'EMBASE'

1683780 2001-2004/PY

L42 21 L30 NOT 2001-2004/PY

FILE 'HCAPLUS'

3686779 2001-2004/PY

L43 48 L31 NOT 2001-2004/PY

FILE 'NTIS'

52701 2001-2004/PY

L44 0 L32 NOT 2001-2004/PY

FILE 'ESBIOBASE'

1050648 2001-2004/PY

L45 12 L33 NOT 2001-2004/PY

FILE 'BIOTECHNO'

368875 2001-2004/PY

L46 19 L34 NOT 2001-2004/PY

FILE 'WPIDS'

3446783 2001-2004/PY

L47 3 L35 NOT 2001-2004/PY

TOTAL FOR ALL FILES

L48 210 L36 NOT 2001-2004/PY

=> s (transcription factor# or splice or poly(w)'a' or polyadenylat? or promoter) (5a) (site# or sequence#) (15a) (reduc? or lower? or decreas?)

FILE 'MEDLINE'
 226841 TRANSCRIPTION
 2158523 FACTOR#
 91045 TRANSCRIPTION FACTOR#
 (TRANSCRIPTION (W) FACTOR#)
 12312 SPLICE
 54177 POLY
 7807314 'A'
 6670 POLYADENYLAT?
 102361 PROMOTER
 657437 SITE#
 706631 SEQUENCE#
1129262 REDUC?
 634492 LOWER?
 908489 DECREAS?
L49 983 (TRANSCRIPTION FACTOR# OR SPLICE OR POLY(W) 'A' OR POLYADENYLAT?
 OR PROMOTER) (5A) (SITE# OR SEQUENCE#) (15A) (REDUC? OR LOWER? OR
 DECREAS?)

FILE 'SCISEARCH'
 180008 TRANSCRIPTION
 1264388 FACTOR#
 68513 TRANSCRIPTION FACTOR#
 (TRANSCRIPTION (W) FACTOR#)
 13470 SPLICE
 153612 POLY
 9624103 'A'
 5237 POLYADENYLAT?
 102707 PROMOTER
 689078 SITE#
 566283 SEQUENCE#
1267124 REDUC?
 712992 LOWER?
 890733 DECREAS?
L50 1233 (TRANSCRIPTION FACTOR# OR SPLICE OR POLY(W) 'A' OR POLYADENYLAT?
 OR PROMOTER) (5A) (SITE# OR SEQUENCE#) (15A) (REDUC? OR LOWER? OR
 DECREAS?)

FILE 'LIFESCI'
 95466 "TRANSCRIPTION"
 289485 FACTOR#
 33534 TRANSCRIPTION FACTOR#
 ("TRANSCRIPTION" (W) FACTOR#)
 6312 SPLICE
 17183 POLY
2030127 'A'
 4136 POLYADENYLAT?
 56650 PROMOTER
 258037 SITE#
 259735 SEQUENCE#
282558 REDUC?
 138702 LOWER?
 216684 DECREAS?
L51 905 (TRANSCRIPTION FACTOR# OR SPLICE OR POLY(W) 'A' OR POLYADENYLAT?
 OR PROMOTER) (5A) (SITE# OR SEQUENCE#) (15A) (REDUC? OR LOWER? OR
 DECREAS?)

FILE 'BIOTECHDS'
 14230 TRANSCRIPTION
 34606 FACTOR#
 1793 TRANSCRIPTION FACTOR#
 (TRANSCRIPTION (W) FACTOR#)
 1206 SPLICE
 6358 POLY

323006 'A'
1530 POLYADENYLAT?
30245 PROMOTER
33811 SITE#
98190 SEQUENCE#
44846 REDUC?
16363 LOWER?
21542 DECREAS?
L52 170 (TRANSCRIPTION FACTOR# OR SPLICE OR POLY(W) 'A' OR POLYADENYLAT?
OR PROMOTER) (5A) (SITE# OR SEQUENCE#) (15A) (REDUC? OR LOWER? OR
DECREAS?)

FILE 'BIOSIS'

205991 TRANSCRIPTION
1207743 FACTOR#
62046 TRANSCRIPTION FACTOR#
(TRANSCRIPTION (W) FACTOR#)
13250 SPLICE
139273 POLY
7719073 'A'
7527 POLYADENYLAT?
111949 PROMOTER
677711 SITE#
519326 SEQUENCE#
1182682 REDUC?
703923 LOWER?
1026285 DECREAS?
L53 994 (TRANSCRIPTION FACTOR# OR SPLICE OR POLY(W) 'A' OR POLYADENYLAT?
OR PROMOTER) (5A) (SITE# OR SEQUENCE#) (15A) (REDUC? OR LOWER? OR
DECREAS?)

FILE 'EMBASE'

210245 "TRANSCRIPTION"
1108877 FACTOR#
60798 TRANSCRIPTION FACTOR#
("TRANSCRIPTION" (W) FACTOR#)
10885 SPLICE
47033 POLY
6772410 'A'
6883 POLYADENYLAT?
87509 PROMOTER
544809 SITE#
495365 SEQUENCE#
1065611 REDUC?
588668 LOWER?
854645 DECREAS?
L54 1303 (TRANSCRIPTION FACTOR# OR SPLICE OR POLY(W) 'A' OR POLYADENYLAT?
OR PROMOTER) (5A) (SITE# OR SEQUENCE#) (15A) (REDUC? OR LOWER? OR
DECREAS?)

FILE 'HCAPLUS'

256737 TRANSCRIPTION
1360433 FACTOR#
119085 TRANSCRIPTION FACTOR#
(TRANSCRIPTION (W) FACTOR#)
15285 SPLICE
615264 POLY
18101164 'A'
10741 POLYADENYLAT?
150102 PROMOTER
852424 SITE#
724307 SEQUENCE#
1851950 REDUC?
819040 REDN

2302369 REDUC?
 (REDUC? OR REDN)
1289399 LOWER?
2093611 DECREAS?
L55 1717 (TRANSCRIPTION FACTOR# OR SPLICE OR POLY(W) 'A' OR POLYADENYLAT?
 OR PROMOTER) (5A) (SITE# OR SEQUENCE#) (15A) (REDUC? OR LOWER? OR
 DECREAS?)

FILE 'NTIS'
1687 TRANSCRIPTION
146390 FACTOR#
 372 TRANSCRIPTION FACTOR#
 (TRANSCRIPTION(W) FACTOR#)
 460 SPLICE
 5560 POLY
1649160 'A'
 10 POLYADENYLAT?
 921 PROMOTER
121027 SITE#
 28227 SEQUENCE#
177425 REDUC?
 66601 LOWER?
 51141 DECREAS?
L56 0 (TRANSCRIPTION FACTOR# OR SPLICE OR POLY(W) 'A' OR POLYADENYLAT?
 OR PROMOTER) (5A) (SITE# OR SEQUENCE#) (15A) (REDUC? OR LOWER? OR
 DECREAS?)

FILE 'ESBIOBASE'
103054 TRANSCRIPTION
399685 FACTOR#
 45149 TRANSCRIPTION FACTOR#
 (TRANSCRIPTION(W) FACTOR#)
 7894 SPLICE
 15825 POLY
2133392 'A'
 2581 POLYADENYLAT?
 54889 PROMOTER
421651 SITE#
 227247 SEQUENCE#
390336 REDUC?
 213986 LOWER?
 310721 DECREAS?
L57 942 (TRANSCRIPTION FACTOR# OR SPLICE OR POLY(W) 'A' OR POLYADENYLAT?
 OR PROMOTER) (5A) (SITE# OR SEQUENCE#) (15A) (REDUC? OR LOWER? OR
 DECREAS?)

FILE 'BIOTECHNO'
160885 TRANSCRIPTION
296524 FACTOR#
 41412 TRANSCRIPTION FACTOR#
 (TRANSCRIPTION(W) FACTOR#)
 8894 SPLICE
 21682 POLY
1454372 'A'
 5860 POLYADENYLAT?
 72959 PROMOTER
222731 SITE#
 375038 SEQUENCE#
232937 REDUC?
 106436 LOWER?
 171676 DECREAS?
L58 1101 (TRANSCRIPTION FACTOR# OR SPLICE OR POLY(W) 'A' OR POLYADENYLAT?
 OR PROMOTER) (5A) (SITE# OR SEQUENCE#) (15A) (REDUC? OR LOWER? OR
 DECREAS?)

FILE 'WPIDS'
 12170 TRANSCRIPTION
 148873 FACTOR#
 1897 TRANSCRIPTION FACTOR#
 (TRANSCRIPTION(W) FACTOR#)
 9528 SPLICE
 155046 POLY
 1692882 'A'
 863 POLYADENYLAT?
 30580 PROMOTER
 115016 SITE#
 233214 SEQUENCE#
1984326 REDUC?
 61106 REDN
2009742 REDUC?
 (REDUC? OR REDN)
1139302 LOWER?
204740 DECREAS?
L59 131 (TRANSCRIPTION FACTOR# OR SPLICE OR POLY(W) 'A' OR POLYADENYLAT?
 OR PROMOTER) (5A) (SITE# OR SEQUENCE#) (15A) (REDUC? OR LOWER? OR
 DECREAS?)

TOTAL FOR ALL FILES

L60 9479 (TRANSCRIPTION FACTOR# OR SPLICE OR POLY(W) 'A' OR POLYADENYLAT?
 OR PROMOTER) (5A) (SITE# OR SEQUENCE#) (15A) (REDUC? OR LOWER? OR
 DECREAS?)

=> s l12 and 160

FILE 'MEDLINE'

L61 72 L1 AND L49

FILE 'SCISEARCH'

L62 67 L2 AND L50

FILE 'LIFESCI'

L63 52 L3 AND L51

FILE 'BIOTECHDS'

L64 38 L4 AND L52

FILE 'BIOSIS'

L65 70 L5 AND L53

FILE 'EMBASE'

L66 79 L6 AND L54

FILE 'HCAPLUS'

L67 158 L7 AND L55

FILE 'NTIS'

L68 0 L8 AND L56

FILE 'ESBIOBASE'

L69 50 L9 AND L57

FILE 'BIOTECHNO'

L70 67 L10 AND L58

FILE 'WPIDS'

L71 44 L11 AND L59

TOTAL FOR ALL FILES

L72 697 L12 AND L60

=> s 112(15a)160
FILE 'MEDLINE'
L73 14 L1 (15A)L49

FILE 'SCISEARCH'
L74 15 L2 (15A)L50

FILE 'LIFESCI'
L75 12 L3 (15A)L51

FILE 'BIOTECHDS'
L76 7 L4 (15A)L52

FILE 'BIOSIS'
L77 11 L5 (15A)L53

FILE 'EMBASE'
L78 21 L6 (15A)L54

FILE 'HCAPLUS'
L79 59 L7 (15A)L55

FILE 'NTIS'
L80 0 L8 (15A)L56

FILE 'ESBIOBASE'
L81 12 L9 (15A)L57

FILE 'BIOTECHNO'
L82 14 L10(15A)L58

FILE 'WPIDS'
L83 15 L11(15A)L59

TOTAL FOR ALL FILES
L84 180 L12(15A) L60

=> s 124 and 160
FILE 'MEDLINE'
L85 0 L13 AND L49

FILE 'SCISEARCH'
L86 0 L14 AND L50

FILE 'LIFESCI'
L87 0 L15 AND L51

FILE 'BIOTECHDS'
L88 0 L16 AND L52

FILE 'BIOSIS'
L89 0 L17 AND L53

FILE 'EMBASE'
L90 0 L18 AND L54

FILE 'HCAPLUS'
L91 1 L19 AND L55

FILE 'NTIS'
L92 0 L20 AND L56

FILE 'ESBIOBASE'

L93 0 L21 AND L57

FILE 'BIOTECHNO'

L94 0 L22 AND L58

FILE 'WPIDS'

L95 0 L23 AND L59

TOTAL FOR ALL FILES

L96 1 L24 AND L60

=> s (l84 or l96) not 2001-2004/py

FILE 'MEDLINE'

1967013 2001-2004/PY

L97 11 (L73 OR L85) NOT 2001-2004/PY

FILE 'SCISEARCH'

3631421 2001-2004/PY

L98 11 (L74 OR L86) NOT 2001-2004/PY

FILE 'LIFESCI'

359020 2001-2004/PY

L99 12 (L75 OR L87) NOT 2001-2004/PY

FILE 'BIOTECHDS'

76520 2001-2004/PY

L100 4 (L76 OR L88) NOT 2001-2004/PY

FILE 'BIOSIS'

1871918 2001-2004/PY

L101 9 (L77 OR L89) NOT 2001-2004/PY

FILE 'EMBASE'

1683780 2001-2004/PY

L102 16 (L78 OR L90) NOT 2001-2004/PY

FILE 'HCAPLUS'

3686779 2001-2004/PY

L103 21 (L79 OR L91) NOT 2001-2004/PY

FILE 'NTIS'

52701 2001-2004/PY

L104 0 (L80 OR L92) NOT 2001-2004/PY

FILE 'ESBIOBASE'

1050648 2001-2004/PY

L105 8 (L81 OR L93) NOT 2001-2004/PY

FILE 'BIOTECHNO'

368875 2001-2004/PY

L106 14 (L82 OR L94) NOT 2001-2004/PY

FILE 'WPIDS'

3446783 2001-2004/PY

L107 3 (L83 OR L95) NOT 2001-2004/PY

TOTAL FOR ALL FILES

L108 109 (L84 OR L96) NOT 2001-2004/PY

=> dup rem 148,1108

PROCESSING COMPLETED FOR L48

PROCESSING COMPLETED FOR L108

L109 105 DUP REM L48 L108 (214 DUPLICATES REMOVED)

=> d tot

L109 ANSWER 1 OF 105 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN
TI Hygromycin-tolerant **gene** with CTG codon **modified** into
leucine **codon**, applicable as **selection** marker in
yeast of Candida genus providing transformants for efficient production
of e.g. dicarboxylic acid;
plasmid pUCARS-HGM-mediated gene transfer and expression in Candida
tropicalis

AU Tanaka A; Ueda M; Hara A; Misawa A
AN 2001-04352 BIOTECHDS
PI WO 2000075307 14 Dec 2000

L109 ANSWER 2 OF 105 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN
TI Constructing **synthetic** polynucleotide for targeting expression
of **gene** to particular cells or tissues, involves substituting
one or more codons or parent polynucleotide encoding protein with a
synonymous codon;

plasmid pAOV2-mediated gene transfer and expression in Escherichia
coli or transgenic plant using Agrobacterium sp. for gene targeting

AU Zhou J; Frazer I H; Botella Mesa J R
AN 2000-12546 BIOTECHDS
PI WO 2000042190 20 Jul 2000

L109 ANSWER 3 OF 105 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN
TI Genotype analysis method, defined as SOMA (short oligonucleotide mass
analysis), of short, defined amplification products using electro-spray
ionization mass spectrometry, useful for analyzing the genotype of living
organisms;

for human genotyping and polymorphism detection using DNA primer
AU Laken S J; Vogelstein B; Kinzler K W; Groopman J D; Jackson P E; Friesen
M D
AN 2000-11281 BIOTECHDS
PI WO 2000031300 2 Jun 2000

L109 ANSWER 4 OF 105 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN
TI **Modified synthetic DNA sequences** comprise
modification of the truncated cry9Aa **gene** of Bacillus
thuringiensis for improved insect control in plants;

transgenic plant construction with improved disease-resistance
AU Kuvshinov V; Kanerva A; Koivu K; Pehu E
AN 2000-06780 BIOTECHDS
PI WO 2000011025 2 Mar 2000

L109 ANSWER 5 OF 105 HCPLUS COPYRIGHT 2004 ACS on STN
TI Recombinant bioadhesive protein analogs comprising hydroxyproline
SO PCT Int. Appl., 52 pp.
CODEN: PIXXD2

IN Paolella, David N.; Gruskin, Elliott A.; Buechter, Douglas D.
AN 2000:191212 HCPLUS
DN 132:232726

| PATENT NO. | KIND | DATE | APPLICATION NO. | DATE |
|---------------|---|----------|-----------------|----------|
| ----- | ----- | ----- | ----- | ----- |
| WO 2000015789 | A1 | 20000323 | WO 1999-US20463 | 19990907 |
| W: | AL, AM, AT, AU, AZ, BB, BG, BR, BY, CA, CH, CN, CZ, DE, DK, EE,
ES, FI, GB, GE, HU, IL, IS, JP, KE, KG, KP, KR, KZ, LK, LR, LS,
LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD,
SE, SG, SI, SK, TJ, TM, TR, TT, UA, UG, UZ, VN | | | |
| RW: | AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL,
PT, SE | | | |
| AU 9959100 | A1 | 20000403 | AU 1999-59100 | 19990907 |

L109 ANSWER 6 OF 105 HCPLUS COPYRIGHT 2004 ACS on STN
TI Molecular characterization of Drosophila melanogaster dihydropteridine

- reductase
- SO Biochimica et Biophysica Acta (2000), 1492(1), 247-251
CODEN: BBACAO; ISSN: 0006-3002
- AU Park, Dongkook; Park, Sangick; Yim, Jeongbin
AN 2000:504957 HCAPLUS
DN 133:219415
- L109 ANSWER 7 OF 105 HCAPLUS COPYRIGHT 2004 ACS on STN
TI Design and cloning of a **modified synthetic gene** for flounder antifreeze peptide
- SO Neimenggu Daxue Xuebao, Ziran Kexueban (2000), 31(2), 216-222
CODEN: NDZKEJ; ISSN: 1000-1638
- AU Erden-Dalai, Wu, Zhu, Ye-rong; Ma, Zhen-yi; Wang, Fei; Kan, Rui
AN 2000:307659 HCAPLUS
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L109 ANSWER 103 OF 105 HCAPLUS COPYRIGHT 2004 ACS on STN

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L109 ANSWER 104 OF 105 WPIDS COPYRIGHT 2004 THOMSON DERWENT on STN

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L109 ANSWER 105 OF 105 MEDLINE on STN DUPLICATE 51

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L109 ANSWER 2 OF 105 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN

AB Constructing a synthetic polynucleotide which selectively expresses a protein in a target cell from plant, relative to another cell from the plant by selecting a first codon of parent polynucleotide and replacing it with synonymous codon which has a higher translational efficiency in the target cell than the other cell, is new. Also claimed are: a polynucleotide (762 or 780 bp) constructed by the method; a vector (e.g. plasmid pAOV2); a cell (e.g. Escherichia coli) containing the polynucleotide; a cell containing the vector; a transgenic plant or its parts produced using Agrobacterium sp.-mediated transfer; selectively expressing a protein in a target plant cell; expressing a

protein from a first nucleotide in a target plant cell; and a cell produced by the method. To facilitate selective expression of proteins to a particular target plant cell or tissue and target expression to particular cells or tissues to produce transgenic plants with novel phenotypes e.g. conferring herbicides the resistance in leaves of a plant but not in the roots of the plant. (106pp)

L109 ANSWER 4 OF 105 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN
AB A **modified synthetic DNA sequence** (I) for improved insect control comprising DNA modified from the truncated crystal protein cry9Aa gene of *Bacillus thuringiensis* ssp. *galleriae* are claimed. The encoded crystal protein has a disclosed protein sequence (Ia) of 624 amino acid residues or has a **sequence alteration**, but the same activity as the active N-terminal domain of the Cry9Aa protein. Also claimed are: a DNA construct for cloning and/or transforming prokaryotic or eukaryotic organisms comprising (I); a prokaryotic or eukaryotic host comprising (I); a method for preparing (I) involving selecting a DNA sequence encoding (Ia) and the unique properties of the Cry9Aa protein and differing from other CryI proteins, providing **synthetic DNA sequences** encoding (Ia), which is encoded by the truncated DNA sequence of 1,989 bp obtainable from the native cry9Aa gene having a 3,837 bp sequence by trypsin (EC-3.4.21.4) cleavage, and improving translation by changing the **codon preference**; and a method for providing higher plants with insect resistance involving incorporating (I) into a DNA construct and incorporating the construct into a plant to give a transgenic plant. (90pp)

L109 ANSWER 7 OF 105 HCPLUS COPYRIGHT 2004 ACS on STN
AB Codon usage varies throughout the animal and plant kingdoms. The flounder antifreeze peptide (AFP) is Ala-rich (60%) and the Ala codons of the fish cDNA reveal a strong bias toward GCC (23/38). We have taken the opportunity to modify the **codon selection** for all the amino acids to be consistent with normal usage in plant genes. The pre-sequence of AFP was replaced by the pre- sequence of sweet potato Sporamin A (EMBL X13 509), and the pro sequence was not included. The **modified genes** coding 2,4,8 tandem mature AFP were constructed and confirmed by DNA sequencing. Using QIA express system. expression of DHFR-mature AFP constructs in *E. coli* suggests the **modified synthetic AFP gene** could be work in plants.

L109 ANSWER 10 OF 105 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN
AB Molecular biology of spider silk was studied with respect to silk processing, genetic engineering and potential application. In vitro production of silk from excised major ampullate gland was demonstrated. A total of 2.3 kb was sequenced. cDNA clones of greater than 2.3 kb for the major ampullate silk gene was unstable, gene **codon preferences** and incompatibility with prokaryote hosts such as *Escherichia coli* was speculated as a problem due to mismatched tRNA availability. The use of **synthetic genes**, constructed from small size oligonucleotide repeats allowed control over primary sequence and final protein size. Expression levels from the **synthetic genes** were low and generally represented up to 5% of the total protein in the cell. Yield were low and generally in the 1-10 mg/l range, depending on the size of the protein. Additional research directions for bioengineering spider silk proteins e.g. fibroin include transgenic expression in the silkworm as well as other organisms as a route to improving yields of the materials in order to explore and extend the potential applications for these materials. (60 ref)

L109 ANSWER 11 OF 105 MEDLINE on STN DUPLICATE 3
AB The catalytic domain of the xynB (xylanase) gene from the thermophilic bacterium *Dictyoglomus thermophilum* was reconstructed by PCR to match the

codon preference of *Trichoderma reesei*. The 0.6-kb DNA fragment encoding the enzyme was first amplified by primer extension with a mixture of eight overlapping oligonucleotides, followed by PCR with outside primers containing restriction enzyme sites for directional cloning into *Escherichia coli* and *T. reesei* vectors. The **synthetic gene** was expressed in both organisms, producing a clearing halo around transformant colonies in plate assay utilizing an overlay of oat spelt xylan. Effective transcription of *xyn B* in *T. reesei* was obtained after changing 20 codons.

- L109 ANSWER 12 OF 105 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN
AB A DNA molecule (I) encoding a dehalogenase that is capable of dalapon herbicide pesticide degradation is claimed, where the DNA has codon usage suitable for small grain species. Also claimed are: a vector which contains (I); an expression cassette containing (I) or a vector, which is suitable for the transformation of plant cells and plants of small grain species and to confer dalapon-resistance to regenerated plants; a transgenic plant of a small grain species containing (I) and that is substantially resistant to dalapon at field use levels; seeds and plants of small grain species which possess, stably integrated in their genome, a foreign DNA such as (I) sufficient to render the small grain species resistant to dalapon at field use levels; a method for rendering a plant of a small grain species resistant to dalapon at field use levels; and a method for protecting plants of a small grain species and destroying weeds in a field using the herbicide dalapon. (I) is *Pseudomonas* sp. **dehal gene** with its **codon preference altered**. The plants are preferably wheat (*Triticum aestivum*) and exhibit herbicide resistance. (29pp)

- L109 ANSWER 13 OF 105 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN
AB A means of transforming a higher plant with a foreign gene containing a modified polyadenylation signal sequence, is claimed. Also claimed is a plant transformed in this manner, the seed of that plant, and nucleic acids containing the modified gene sequence. This is used to produce transgenic plants that produce an increased amount of a desired gene, particularly to improve the iron absorption efficiency of crop plants. The **modified gene** preferably encodes a protein involved in nutrient absorption, such as a ferric iron-**reductase** gene derived from yeast. The **polyadenylation sequence** is **modified** by base substitution, based on the codon use ratio of the plant to be transformed, preferably so that the modified codons reduce the same amino acid residues as the original. The foreign gene preferably includes a Kozak initiation sequence upstream of the start codon of the gene. This is particularly used in the production of transgenic tobacco (*Nicotiana tabacum*) and transgenic rice (*Oryza sativa*). The vector is specifically vector plasmid pBI121, which is transferred into plant cells using *Agrobacterium tumefaciens*. (81pp)

- L109 ANSWER 19 OF 105 MEDLINE on STN DUPLICATE 4
AB The *Plasmodium falciparum* malaria parasite is the causative agent of malaria tropica. Merozoites, one of the extracellular developmental stages of this parasite, expose at their surface the merozoite surface protein-1 complex (MSP-1), which results from the proteolytic processing of a 190-200 kDa precursor. MSP-1 is highly immunogenic in humans and numerous studies suggest that this protein is an effective target for a protective immune response. Although its function is unknown, there are indications that it may play a role during invasion of erythrocytes by merozoites. The parasite-derived *msp-1* gene, which is approximately 5000 bp long, contains 74% AT. This high AT content has prevented stable cloning of the full-size gene in *Escherichia coli* and consequently its expression in heterologous systems. Here, we describe the synthesis of a 4917 bp gene encoding MSP-1 from the FCB-1 strain of *P. falciparum* adjusted for human **codon preferences**. The **synthetic msp-1 gene** (55% AT) was cloned, maintained and

expressed in its entirety in E.coli as well as in CHO and HeLa cells. The purified protein is soluble and appears to possess native conformation because it reacts with a panel of mAbs specific for conformational epitopes. The strategy we used for synthesizing the full-length msp-1 gene was to assemble it from DNA fragments encoding all of the major proteolytic fragments normally generated at the parasite's surface. Thus, after subcloning we also obtained each of these MSP-1 processing products as hexahistidine fusion proteins in E.coli and isolated them by affinity chromatography on Ni²⁺-agarose. The availability of defined preparations of MSP-1 and its major processing products open up new possibilities for in-depth studies at the structural and functional level of this important protein, including the exploration of MSP-1-based experimental vaccines.

L109 ANSWER 21 OF 105 HCAPLUS COPYRIGHT 2004 ACS on STN

AB The invention provides a compound comprising a nucleic acid encoding an 85 antigen of Mycobacterium, wherein at least one codon of the **nucleic acid** encoding the 85 antigen is **altered** whereby expression of the 85 antigen in an expression system is increased over expression in the same expression system of a nucleic acid having a wild-type codon at the same position as the altered codon. Specifically, the invention provides an isolated nucleic acid encoding a protein of Mycobacterium comprising at least one codon **altered** to increase expression of the **nucleic acid**, wherein the **codon** is **selected** from the group consisting of AGG, AGA, ATA, CGA, CTA, CGG, CTT, CTC, GGG, and GGA and an isolated nucleic acid encoding an 85 antigen of Mycobacterium comprising at least one codon **altered** to increase expression of the **nucleic acid**, wherein the **codon** is **selected** from the group consisting of AGG, AGA, ATA, CTA, CGC, CTT, CTC, GGG, and GGA. Also provided is an improved method of producing a Mycobacterium protein in a host cell comprising **altering** a codon of a **nucleic acid** encoding the Mycobacterium protein so that the **altered** codon of the **nucleic acid** is one preferred by the host and introducing the **nucleic acid** containing the **altered** codon into the host, whereby the host expresses the nucleic acid thereby producing the Mycobacterium protein. When the wild-type antigen 85C gene was expressed in E. coli, the yield was 15-20 mg antigen/L. When five codons in this gene were changed to E. coli-preferred codons, the yield increased to >60 mg/L.

L109 ANSWER 27 OF 105 MEDLINE on STN

DUPLICATE 7

AB **Synthetic genes** are very useful in genetic and protein engineering. Here we propose a general method for construction of **synthetic genes**. Short oligonucleotides are joined through ligase chain reaction (LCR) in high stringency conditions to make "unit fragments" which are then fused to form a full-length gene sequence by polymerase chain reaction. The procedure is simple and accurate and does not place constraints on sequence and length. In this report, a recombinant leptin gene was synthesized according to the **codon preference** of Escherichia coli. Besides, a substitution of the only Met at position 54 for Leu and an addition of a Met at the N-terminus were introduced in the **synthetic gene**. The gene was cloned in the pQE-31 expression vector and was expressed in E. coli. A large amount of recombinant leptin containing 6 x His tag was produced and purified by Ni-NTA affinity column. Finally, intact leptin-L54 was released after removing the tag by CNBr cleavage at the Met residue.

L109 ANSWER 30 OF 105 HCAPLUS COPYRIGHT 2004 ACS on STN

AB A novel cecropin AD gene was designed and synthesized, in which DNA sequence is based on the amino acid sequence of hybrid peptide cecropin A1-11D12-37. The **codon selection** of the designed gene was carried out according to the codon usage of yeast. The designed gene is 140 bp in length, including the encoded sequence, the start and stop codons, and the restriction sites of BamH I, EcoR I and Sal I at both

ends. The cecropin AD gene was synthesized by twice PCR method and cloned into PCRTM 2.1 vector. It was shown that the DNA **sequence** of the **synthetic** cecropin AD **gene** coincides with that of the designed gene by DNA sequencing.

L109 ANSWER 33 OF 105 HCAPLUS COPYRIGHT 2004 ACS on STN

AB A review with 60 refs. of an ongoing process to genetically engineer durable pathogen resistance into poplar and other tree species. The process involved designing, testing, and selecting antimicrobial peptide sequences to use as transgenic plant products. The **synthetic** peptide **sequence** were then encoded into DNA. Gene promoters, translation initiation and termination sequences, **codon preferences**, mRNA secondary structure, peptide stability and peptide targeting were considered when designing gene constructs to test in transgenic plants.

L109 ANSWER 36 OF 105 MEDLINE on STN DUPLICATE 12

AB **Synthetic genes** were designed to encode analogs of the two proteins of Nephila clavipes dragline silk, spidroins 1 and 2. The genes were constructed of tandem repeats of relatively long (more than 300 bp) DNA **sequences** assembled from **synthetic** oligonucleotides, and encoded proteins of high molecular mass (65-163 kDa). Both analogs were produced efficiently in Escherichia coli. The yield and homogeneity of the products of longer genes were limited by premature termination of synthesis, probably as a result of processivity errors in protein synthesis. Average termination rates were determined to be 1 in 1100 codons to 1 in 300 codons, depending on the length and synonymous **codon choices** of the gene. Both analog proteins could be induced to form stable aqueous solutions without denaturants. Circular dichroism spectra of the purified proteins in dilute solution resembled spectra of redissolved natural dragline silk in reflecting a largely disordered structure in water and more ordered structures in mixed solvents with methanol and trifluoroethanol.

L109 ANSWER 41 OF 105 MEDLINE on STN DUPLICATE 15

AB BACKGROUND. The expression of both the env and gag gene products of human immunodeficiency virus type 1 (HIV-1) is known to be limited by cis elements in the viral RNA that impede egress from the nucleus and reduce the efficiency of translation. Identifying these elements has proven difficult, as they appear to be disseminated throughout the viral genome. RESULTS. Here, we report that **selective codon usage** appears to account for a substantial fraction of the inefficiency of viral protein synthesis, independent of any effect on improved nuclear export. The codon usage effect is not specific to transcripts of HIV-1 origin. Re-engineering the coding sequence of a model protein (Thy-1) with the most prevalent HIV-1 codons significantly impairs Thy-1 expression, whereas **altering** the coding **sequence** of the jellyfish green fluorescent protein gene to conform to the favored codons of highly expressed human proteins results in a substantial increase in expression efficiency. CONCLUSIONS. Codon-usage effects are a major impediment to the efficient expression of HIV-1 genes. Although mammalian genes do not show as profound a bias as do Escherichia coli genes, other proteins that are poorly expressed in mammalian cells can benefit from codon re-engineering.

L109 ANSWER 42 OF 105 LIFESCI COPYRIGHT 2004 CSA on STN DUPLICATE 16

AB The cloning and expression of Bacillus thuringiensis delta -endotoxin genes in transgenic plants have been used with the objective of protecting the crops from insect attack. The increased expression of the insecticidal cry genes in plants has been critical for the development of genetically transformed plants with agronomically acceptable levels of insect resistance. Low expression levels of such genes also have an environmental implication: the release of low expressing insect-tolerant transgenic plants may result in the rapid appearance of resistance to the Cry toxin

in the target insect. The problem of the expression of *B. thuringiensis* cry genes is due to the expression of bacterial prokaryotic genes in higher plants or in any other eukaryotic organism. Fully **modified genes** can express up to 100-fold higher levels of the insecticidal toxin compared to those obtained when a wild-type bacterial gene is expressed. We describe the most important aspects present in the bacterial wild-type cry genes affecting their expression in transgenic plants. The analysis includes aspects of transcriptional regulation, mRNA stability, **preferences** in codon usage and translational efficiency. According to these considerations, **modified cry genes** have been reconstructed allowing to increase the expression levels in transgenic plants.

L109 ANSWER 44 OF 105 MEDLINE on STN DUPLICATE 18
AB Considering the factors which affect gene transcription, translation and the stability of mRNA, without changing the amino acid composition of the encoded polypeptide, AaIT gene encoding insect-specific neurotoxin was designed and synthesized according to bias in **codon choice**, overall G+C content and G+C content of bases at the third position in codons of polyhedrin genes of baculovirus and of plant genes as well. AaIT **gene** was fused behind a **synthetic gp67 signal sequence** and then recombined into the genome of *Trichoplusia ni* nuclear polyhedrosis virus (TnNPV) by transfer vector pSXIV VI+X3. The recombinant virus TnNPV-AaIT (occ+-gal-) was screened. The results of Southern blotting and SDS-PAGE demonstrated that AaIT gene had integrated into the genome of virus and expressed. Bioassays on the 3rd-instar *Trichoplusia ni* larvae showed that recombinant viruses TnNPV-AaIT could shorten the time of killing insect and improve the efficacy of killing agronomically important insects.

L109 ANSWER 46 OF 105 MEDLINE on STN DUPLICATE 20
AB Plasmodium falciparum dihydrofolate reductase-thymidylate synthase (DHFR-TS) is a well-known target for pyrimethamine and cycloguanil. The low amounts of enzyme obtainable from parasites or the currently available heterologous expression systems have thus far hindered studies of this enzyme. The 1912-base pair *P. falciparum* DHFR-TS gene was designed based on *E. coli* **codon preference** with unique restriction sites evenly placed throughout the coding sequence. The gene was designed and synthesized as three separated domains: the DHFR domain, the junctional sequence, and the TS domain. Each of these domains contained numerous unique restriction sites to facilitate mutagenesis. The three domains were assembled into a complete DHFR-TS gene which contained 30 unique restriction sites in the coding sequence. The bifunctional DHFR-TS was expressed from the **synthetic gene** as soluble enzyme in *E. coli* about 10-fold more efficiently than from the wild-type sequence. The DHFR-TS from the **synthetic gene** had kinetic properties similar to those of the wild-type enzyme and represents a convenient source of protein for further study. The unique restriction sites in the coding sequence permits easy mutagenesis of the gene which should facilitate further understanding of the molecular basis of antifolate resistance in malaria.

L109 ANSWER 47 OF 105 MEDLINE on STN DUPLICATE 21
AB A synthetic wheat high-molecular-weight (HMW) glutenin storage protein gene analog was constructed for expression in *E. coli*. This first **synthetic HMW-glutenin gene** and future **modifications** are intended to allow systematic dissection of the molecular basis of HMW-glutenin role in the visco-elastic properties critical for wheat product processing and utilization. The design of the gene included four features: different construction strategies for the separate assembly of major polypeptide domains, the inclusion of convenient restriction sites for modifications, use of a **codon selection** similar to *E. coli* highly expressed genes, and the ability to produce repetitive sequence domains of exact numbers of defined

repeats. The complete synthetic HMW-glutenin construct was 1908 bp, and contained 32 identical copies of one of the HMW-glutenin repetitive domain motifs. The gene expressed the novel HMW-glutenin protein to relatively high levels in bacterial cultures and the protein exhibited the known anomalous behavior of HMW-glutenins in SDS-PAGE.

- L109 ANSWER 49 OF 105 MEDLINE on STN DUPLICATE 22
AB **Synthetic genes** encoding recombinant spider silk proteins have been constructed, cloned, and expressed. Protein sequences were derived from *Nephila clavipes* dragline silk proteins and reverse-translated to the corresponding DNA sequences. **Codon selection** was chosen to maximize expression levels in *Escherichia coli*. DNA "monomer" sequences were multimerized to encode high molecular weight synthetic spider silks using a "head-to-tail" construction strategy. Multimers were cloned into a prokaryotic expression vector and the encoded silk proteins were expressed in *E. coli* upon induction with IPTG. Four multimer, ranging in size from 14.7 to 41.3 kDa, were chosen for detailed analysis. These proteins were isolated by immobilized metal affinity chromatography and purified using reverse-phase HPLC. The composition and identity of the purified proteins were confirmed by amino acid composition analysis, N-terminal sequencing, laser desorption mass spectroscopy, and Western analysis using antibodies reactive to native spider dragline silk. Circular dichroism measurements indicate that the synthetic spider silks have substantial beta-sheet structure.
- L109 ANSWER 54 OF 105 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN
AB Efficient heterologous gene expression during high cell density culture of *Escherichia coli* is critical to the successful commercial application of biotechnology. Various approaches to enhance gene expression, including appropriate promoter **selection**, optimized **codon** usage, ribosome enhancement, host cell genetics **modification**, **gene** dosage effect, plasmid stability, mRNA stability enhancement, host cell genetics modification and amino acid misincorporation control, are discussed in the context of improvement of both recombinant product quality and yield. These approaches will then allow successful fermentation process scale-up. (0 ref)
- L109 ANSWER 56 OF 105 MEDLINE on STN DUPLICATE 27
AB I present evidence that natural **selection** biases synonymous **codon** usage to enhance the accuracy of protein synthesis in *Drosophila melanogaster*. Since the fitness cost of a translational misincorporation will depend on how the amino acid substitution affects protein function, selection for translational accuracy predicts an association between codon usage in DNA and functional constraint at the protein level. The frequency of preferred codons is significantly higher at codons conserved for amino acids than at nonconserved codons in 38 genes compared between *D. melanogaster* and *Drosophila virilis* or *Drosophila pseudoobscura* ($Z = 5.93$, $P < 10(-6)$). Preferred codon usage is also significantly higher in putative zinc-finger and homeodomain regions than in the rest of 28 *D. melanogaster* transcription factor encoding genes ($Z = 8.38$, $P < 10(-6)$). Mutational **alternatives** (within-**gene** differences in mutation rates, amino acid changes altering **codon preference** states, and doublet mutations at adjacent bases) do not appear to explain this association between synonymous codon usage and amino acid constraint.
- L109 ANSWER 57 OF 105 SCISEARCH COPYRIGHT 2004 THOMSON ISI on STN
AB RAP30 and RAP74 are subunits of RAP30/74 (TFIIF, beta gamma), a general initiation and elongation factor for transcription by RNA polymerase II. Methods were previously published for production of human RAP30 and RAP74 in bacterial cells, using a bacteriophage T7 promoter expression system. The vectors described for production of RAP74 were not very efficient and produced significant quantities of RAP74 amino terminal fragments. To

improve these vectors, a segment of the human RAP74 cDNA was recoded using a preferred set of codons for translation in Escherichia coil. Recoding dramatically improved protein production and suppressed production of amino-terminal fragments. Improved vectors are reported that produce RAP74 with an LEHHHHHH carboxy-terminal extension (RAP74-H-6), for purification on a Ni²⁺-affinity column, and also with the native carboxy terminus (RAP74). Methods for purification of RAP74-H-6 and RAP74 are reported. Using these improved vectors, approximately 30 mg of soluble and active RAP74-H-6 or RAP74 can be produced and purified from 1 liter off. coil culture, representing a 10-fold improvement in protein production. Methods have also been developed for reconstitution of native RAP30/74 complex using recombinant proteins. This complex has indistinguishable activity from human RAP30/74 for accurate transcription in vitro. (C) 1994 Academic Press, Inc.

L109 ANSWER 66 OF 105 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN
AB A new method for determining the relative native **codon** pairing **preference** in an organism involves: obtaining nucleotide sequence data; determining the codon usage; determining the expected number of occurrences of randomly-paired codons; comparing the expected number with the actual number to determine the relative codon usage; eliminating any amino acid pair bias; and **altering** a **gene** for expression in the organism, by substituting codons according to the preferred usage, to **alter** the translational kinetics for a **gene** in a predetermined manner. The information may be used for: artificial **gene** construction, e.g. for **altering** a **gene** from a 1st organism for high-level expression in a 2nd organism; to determine the type of organism from which a sample of nucleic acid originates, by comparing the **codon** pair **preference** with standard data; and introducing a translational pause site into a gene, by introduction of an over-represented codon pair. The identification of **codon** pair **preferences** in organisms allows enhanced recombinant protein expression, controlled protein folding and evolutionary studies. (106pp)

L109 ANSWER 67 OF 105 HCAPLUS COPYRIGHT 2004 ACS on STN
AB A computer program, which runs on MS-DOS personal computers, is described that assists in the design of **synthetic genes** coding for proteins. The goal of the program is the design of a gene which (i) contains as many unique restriction sites as possible and (ii) uses a specific codon usage. The gene designed according to the criteria above is (i) suitable for modular mutagenesis expts. and (ii) optimized for expression. The program reverse-translates protein sequences into degenerated DNA sequences, generates a map of potential restriction sites and locates sequence positions where unique restriction sites can be accommodated. The nucleic acid sequence is then refined according to a specific codon usage to remove any degeneration. Unique restriction sites, if potentially present, can be forced into the degenerated nucleic acid sequence by using priority codes assigned to different restriction sequences.

L109 ANSWER 68 OF 105 MEDLINE on STN DUPLICATE 33
AB High levels of active HIV-1 protease (PR) were produced in Escherichia coli, amounting to 8-10% of total cell protein. High production levels were achieved by altering the following parameters: (1) **codon** **preference** of the coding region, (2) A+T-richness at the 5' end of the coding region, and (3) promoter. To circumvent the toxicity of HIV-1 PR in E. coli, the gene was expressed as a fusion protein with two different proteolytic autocleavage sequences. In both the cases, the fusion protein could be cleaved in vivo to give an active molecule with the native sequence at the N terminus.

L109 ANSWER 74 OF 105 MEDLINE on STN DUPLICATE 36
AB A computer program (PINERS) is described for use in the design of

synthetic genes and mixed-probe DNA sequences. A protein sequence is reverse translated with generation of synonymous codons at each position producing a degenerate sequence. In order to locate potential restriction enzyme sites, the degenerate sequence is searched with a library of restriction enzymes for sites that utilize any combination of synonymous codons. These sites are indicated in a map so that they may be incorporated into the **synthetic gene sequence**. The program allows the user to **select** the appropriate **codon** usage table for the organism of interest and then to set a threshold usage frequency below which codons are not generated. PINCERS may also be used to assist in planning the synthesis of mixed-probe DNA sequences for cross-hybridization experiments. It can identify regions of specified length with the protein sequence that have the least overall degeneracy, thereby minimizing the number of probes to be synthesized and, therefore, maximizing the concentration of a given probe sequence.

L109 ANSWER 78 OF 105 HCAPLUS COPYRIGHT 2004 ACS on STN

AB Bovine insulin-like growth factor 2 (bIGF2) was produced in inclusion bodies in the cytoplasm of E. coli and accumulated at high levels: 20-25% of total Coomassie-stained bacterial protein. The level of accumulation of bIGF2 was affected by the choice of codons in the 5' end of the coding sequence and by a rpoH mutation in the host cells. Purified recombinant bIGF2 had the native N terminus and the same mitogenic activity as that of bIGF2 purified from bovine serum.

L109 ANSWER 80 OF 105 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN

AB A computer aided methodology was provided for the generation of artificial genes. This strategy allowed a complete control over factors such as **codon** usage, **selection** for efficient controllable promoters and initiation signals, and enabled elimination of sequences which interfere with efficient transcription or translation (e.g. secondary structures in RNA). Provision of convenient restriction sites for gene manipulation including site-directed mutagenesis for protein engineering were important assets of the technique. The validity of this method was demonstrated by the construction of the human interleukin-2 (IL-2) gene and 3 of its mutants. The artificial genes were expressed in Escherichia coli MC1060 under the control of the E. coli trp promoter/operator. IL-2 expression levels at the stationary phase of bacterial growth were up to 11-15% of total bacterial proteins. The recombinant IL-2 protein precipitated within the bacterial cells as inclusion bodies. IL-2 and its 3 analogs were active in vitro. Similar approaches can be adopted for any other protein of known primary sequence. (14 ref)

L109 ANSWER 92 OF 105 MEDLINE on STN

DUPLICATE 44

AB A DNA duplex coding for the 27 amino acids of secretin has been synthesized and cloned. In designing the sequence of the gene, computer analysis has been applied. The following factors have been considered: **selection of codon** usage in favour of expression in yeast; design of various sites useful in **gene** cloning, **gene modification** and expressed product purification; avoiding the repeat sequences which may interfere in the ligation of the synthetic fragments. The synthesis involved preparation of 12 oligodeoxyribonucleotides (12-mer to 24-mer in length) by phosphate triester and phosphite triester method, purification by polyacrylamide gel electrophoresis (PAGE). A new plasmid pWS1 was constructed by insertion of the enzymatic ligated gene fragment into plasmid pWR13.

L109 ANSWER 96 OF 105 MEDLINE on STN

DUPLICATE 47

AB A region of minimal **codon** degeneracy was **selected** from the amino acid sequence of the amino-terminal alpha I domain of human erythrocyte spectrin to design a 90-base-pair DNA probe for the screening of a human genomic library. Five complementary oligonucleotides were

assembled to form a full-length double-stranded DNA, which was then cloned in an M13 phage vector to generate hybridization probes. Under stringent conditions, a single hybridizing clone was isolated from a total human genomic library. Partial DNA sequence analysis established the 16.8-kilobase-pair isolate as erythrocyte alpha-spectrin by correlation to a known sequence of 131 amino acids. The spectrin 106 amino acid repeat segment is encoded by multiple exons separated by introns of various sizes. Of the 3074 base pairs of DNA sequenced thus far, 12.8% code for amino acids.

- L109 ANSWER 98 OF 105 BIOTECHDS COPYRIGHT 2004 THOMSON DERTWENT/ISI on STN
 AB Through the use of recently developed recombinant DNA methods, high level expression systems for the production of recombinant proteins such as interferons in microorganisms now exist. Using a defined medium and a multiple-phage resistant AM-7 strain of Escherichia coli containing a temperature-sensitive multicopy plasmid under lambda promoter control, a fermentation process has been developed for an analog human interferon-alpha (IFN-alpha) which is a 'consensus' of the known IFN-alpha subtypes (IFN-alpha-Con1). The **gene** for the interferon was **synthetic** and **codon choices** were optimized for expression in E.coli. Cultures were performed at 30 deg in a chemostat, at pH 7 with shaking. Fed-batch culture was also performed. A culture medium containing glucose was used. Yields of up to 7.6×10^{12} U IFN-alpha-Con1/l of fermentation broth were obtained. The combination of a tightly regulated expression system and a controlled feeding schedule was required for the high expression levels and cell densities necessary to give this yield. (20 ref)
- L109 ANSWER 100 OF 105 MEDLINE on STN DUPLICATE 49
 AB A **synthetic** human growth hormone (hGH) **gene** was efficiently expressed under the control of the repressible acid phosphatase promoter in yeast (*Saccharomyces cerevisiae*). More than 10^6 molecules of hormone were formed per cell despite the fact that the gene was constructed with **codon preference** for *Escherichia coli*.
- L109 ANSWER 103 OF 105 HCPLUS COPYRIGHT 2004 ACS on STN
 AB A rapid, efficient procedure for the total (chemical and enzymic) synthesis of linear, double-stranded DNA sequences of up to .apprx.200 base pairs is described. The method involves the chemical synthesis of oligodeoxyribonucleotides with regions of base complementarity, the annealing of the oligonucleotides, and the joining of oligonucleotides by enzymic methods. The **codon preference** of *Escherichia coli* is considered in the synthesis, and the redundancy of the genetic code is exploited to reduce self-complementarity in the oligonucleotides synthesized. The method is employed in the synthesis and cloning of genes for human β -endorphin [61214-51-5] and leucine-5 human β -endorphin [66238-14-0]. Thus, oligodeoxyribonucleotides were chemical synthesized by the described method, which involved a solid support; the oligonucleotides were joined to form genes for human β -endorphin and a leucine-5 derivative. The fusion of genes for β -endorphin and derivs. to genes for β -galactosidase [9031-11-2] or β -lactamase [9073-60-3] was described, as was the cloning of **synthetic genes** in *E. coli*. The preparation of ^{125}I -labeled **synthetic genes** and antibodies to **gene** products was claimed.

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